CHAPTER 17

Idiopathic Giant Cell Myocarditis

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INTRODUCTION

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INTRODUCTION

Idiopathic giant cell myocarditis (IGCM) was first described by Saltykov in a 37-year-old man who died suddenly after surgical drainage of an abscess. Since then, dozens of case reports and a few small series have documented that IGCM is a usually fatal disorder that generally affects young otherwise healthy individuals, although a minority of cases occur in association with autoimmune disorders or thymoma. From 1905 until 1987, all cases were described at autopsy, and survival was generally less than 3 months from symptom onset. In 1993, Ren et al. described prolonged transplant-free survival in 3 patients with IGCM. Several patients whose disease was diagnosed by endomyocardial biopsy survived 1 or more years in association with immunosuppressive treatment. This chapter reviews the pathophysiology, natural history, proposed diagnostic strategy, and treatment options for IGCM.

In the first half of the twentieth century, the term “giant cell myocarditis” was used to describe both granulomatous and diffuse inflammatory myocardial infiltrates that contained multinucleated giant cells. Idiopathic granulomatous myocarditis with giant cells was described in several case reports as cardiac sarcoidosis or giant cell myocarditis. Tesuk first distinguished the well-organized, granulomatous lesions of cardiac sarcoidosis from a diffuse, nongranulomatous infiltrate, which he called giant cell myocarditis. Most authorities since have considered giant cell myocarditis as a distinct clinical and pathologic entity rather than a virulent form of isolated cardiac sarcoidosis. This distinction does not result from a proven mechanistic difference between these disorders. Indeed, about half of the cases of cardiac sarcoidosis are isolated, with no evidence of extracardiac involvement.

IGCM is a pathologic diagnosis (see Fig. 14-15 A and 14-15 B). A diffuse or multifocal inflammatory infiltrate consists of lymphocytes admixed with eosinophils and multinucleated giant cells. Myocyte damage must be evident in association with the inflammatory lesion. Various degrees of fibrosis may be present. Poorly formed granulomas may be seen in giant cell myocarditis, but well-organized follicular granulomas containing central giant cells exclude the diagnosis by definition. The lesions of active lymphocytic myocarditis may occasionally contain an isolated giant cell; nonetheless, giant cell myocarditis can usually be distinguished from lymphocytic myocarditis and granulomatous myocarditis, even on biopsy specimens (Table 17-1).

The differential diagnosis of IGCM includes drug reactions and systemic diseases (Table 17-2). Drug hypersensitivity reaction may manifest as IGCM, with evidence of hypersensitivity in other organs. IGCM has been described after high-dose interleukin-2 treatment for lymphoma, possibly as a result of cytokine imbalance. Giant cell myocarditis has been described in a case of measles myocarditis.

Up to 20% of cases of giant cell myocarditis occur in individuals with other inflammatory or autoimmune disorders, especially inflammatory bowel disease (Table 17-3). Interestingly, a small percentage of patients who present with giant cell myocarditis at
### Table 17-1
Pathologic Findings in Giant Cell Myocarditis, Cardiac Sarcoidosis, and Lymphocytic Myocarditis

<table>
<thead>
<tr>
<th>Diagnosis</th>
<th>Definition</th>
<th>Gross pathology</th>
<th>Microscopic pathology</th>
<th>Comments</th>
</tr>
</thead>
<tbody>
<tr>
<td>Giant cell myocarditis*</td>
<td>Widespread or serpiginous inflammation with myocyte necrosis in the absence of well-formed granulomas or specific etiology</td>
<td>Pale, flabby myocardium; dilation or hypertrophy may be present; treated cases may have extensive scar</td>
<td>Widespread or serpiginous inflammation with giant cells, lymphocytes, and often eosinophils; myocyte necrosis always present; poorly formed granulomas may be seen</td>
<td>See Tables 17-2 and 17-3 and text for differential diagnosis and associated disorders</td>
</tr>
<tr>
<td>Cardiac sarcoidosis†</td>
<td>Granulomatous myocarditis with no evidence of infectious or other specific cause</td>
<td>Sharply defined areas of granulomatous inflammation or scar; preferential involvement of papillary muscles, septum, and base of ventricles</td>
<td>Non-necrotizing granulomas, fibrosis with few eosinophils; myocyte necrosis is rare</td>
<td>Look for other organ involvement, anergy to common antigens, and ACE level to support diagnosis; exclude fungi, mycobacteria, and foreign body reaction with special studies</td>
</tr>
<tr>
<td>Lymphocytic or idiopathic myocarditis‡</td>
<td>A predominantly lymphocytic infiltrate with associated myocyte damage in the absence of acute infarction</td>
<td>Focal or diffuse inflammatory lesions</td>
<td>First pathologic specimen may be active or borderline myocarditis, the latter having no myocyte damage; subsequent samples may be persistent, healing, or healed per “Dallas criteria”</td>
<td>Associated with coxsackie B, adenoviral, and hepatitis C viral infections; biopsy artifacts may resemble myocyte necrosis; rule out many specific causes</td>
</tr>
</tbody>
</table>

ACE, angiotensin-converting enzyme.


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Table 17-2
Disorders That May Resemble Giant Cell Myocarditis

<table>
<thead>
<tr>
<th>Disorder</th>
<th>Associated findings and diagnostic studies</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sarcoidosis</td>
<td>Granulomatous myocarditis without specific cause; fibrosis may be prominent; myocyte necrosis is rare</td>
</tr>
<tr>
<td></td>
<td>Anergy to common antigens; angiotensin-converting enzyme level; look for other organ involvement</td>
</tr>
<tr>
<td>Wegener granulomatosis</td>
<td>Granulomatous myocarditis; renal and upper and lower respiratory disease are generally present; vasculitis may affect the coronary arteries</td>
</tr>
<tr>
<td></td>
<td>c-ANCA, look for classic renal and respiratory findings</td>
</tr>
<tr>
<td>Foreign body reaction</td>
<td>Giant cells may be associated with myocardial reaction to pacemaker leads or ventricular-assist devices</td>
</tr>
<tr>
<td>Hyper-sensitivity myocarditis</td>
<td>Diffuse, primarily interstitial infiltrate with numerous eosinophils; myocyte necrosis and giant cells are infrequent</td>
</tr>
<tr>
<td></td>
<td>Elevated liver function results, eosinophil count, and skin rash</td>
</tr>
<tr>
<td>Cardiac lymphoma</td>
<td>Hodgkin disease rarely reported with myocardial granulomas with giant cells*</td>
</tr>
<tr>
<td></td>
<td>Immunophenotyping, look for extracardiac involvement</td>
</tr>
<tr>
<td>Fungal myocarditis</td>
<td>Coccidioidomycosis, blastomycosis, actinomycosis, and others from granulomas; seen in immunocompromised hosts, often with associated endocarditis or sepsis</td>
</tr>
<tr>
<td></td>
<td>Special stains such as Grocott methenamine silver and periodic acid-Schiff indicated</td>
</tr>
<tr>
<td>Measles</td>
<td>Rare complication of measles; may cause myocyte necrosis and giant cell myocarditis</td>
</tr>
<tr>
<td></td>
<td>Associated clinical findings</td>
</tr>
<tr>
<td>Syphilis</td>
<td>Gumma may rarely involve the myocardium; myocardial involvement is rarely isolated</td>
</tr>
<tr>
<td></td>
<td>Associated clinical findings; VDRL.</td>
</tr>
<tr>
<td>Tuberculosis</td>
<td>Nodular, miliary, and diffuse infiltrative types; myocardial involvement is rarely isolated</td>
</tr>
<tr>
<td></td>
<td>Other organ involvement; PPD skin test</td>
</tr>
<tr>
<td>Rheumatic carditis</td>
<td>Interstitial granulomatous infiltrate without myocyte necrosis in active lesion</td>
</tr>
<tr>
<td></td>
<td>Jones criteria¹</td>
</tr>
</tbody>
</table>

c-ANCA, antineutrophil cytoplasmic antibodies; PPD, purified protein derivative; VDRL, Venereal Disease Research Laboratory.

*Saphir O. Arch Pathol 1942;33:88.

²JAMA 1992;268:2069-2073.

Table 17-3
Disorders Associated With Idiopathic Giant Cell Myocarditis

Inflammatory disorders
Ulcerative colitis\textsuperscript{24,26}
Crohn disease\textsuperscript{15}
Orbital & skeletal myositis\textsuperscript{27}
Myasthenia gravis\textsuperscript{22,28}
Thyroiditis\textsuperscript{13,29,30}
Takayasu arteritis\textsuperscript{31,32}
Rheumatoid arthritis\textsuperscript{33}
Pernicious anemia\textsuperscript{23}
Alopecia totalis vitiligo\textsuperscript{34}

Tumors
Thymoma\textsuperscript{28,35}
Lung carcinoma\textsuperscript{29}
Lymphoma\textsuperscript{36,37}
Sarcoma\textsuperscript{13}

Hypersensitivity reaction
Silicone rubber\textsuperscript{38}
Antiseizure medication\textsuperscript{17}

Miscellaneous
Post-mitral valve surgery\textsuperscript{39}
Mitril stenosis-associated\textsuperscript{40}

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autopsy or explantation have clinically unrecognized granulomatous inflammation in other organs, including the aorta, lungs, liver, and lymph nodes.\textsuperscript{41} These cases suggest that IGCM can be the prime manifestation of a systemic granulomatous process. In our experience, several patients with fatal IGCM have had a granulomatous infiltrate in the lymph nodes or other organs. Therefore, the diagnosis of sarcoidosis or the presence of granulomatous infiltration in other organs does not exclude IGCM.\textsuperscript{13,42}

Disorders that cause myocardial granulomas with associated giant cells may be mistaken for IGCM. Aschoff lesions of rheumatic myocarditis evolve into characteristic, focal interstitial granulomas with giant cells. Tuberculosis and cryptococcosis may also have giant cells within granulomatous lesions.\textsuperscript{43-45} Special stains for organisms should be performed whenever there is a question of infection. Rarely, giant cells may be seen in syphilitic myocarditis.\textsuperscript{45} Foreign body reaction, Wegener granulomatosis,\textsuperscript{36,47} and systemic sarcoidosis must be considered in the differential diagnosis as well. These disorders usually
have distinct clinical presentations and appropriate diagnostic studies usually prevent confusion with IGCM.

The tumor most commonly associated with giant cell myocarditis is thymoma, particularly the spindle cell type.\textsuperscript{13} Myasthenia gravis and myositis are associated with thymoma and giant cell myocarditis individually and in combination.\textsuperscript{20,22,27,31,48-52} The association of thymoma with giant cell myocarditis occurs almost exclusively in women.\textsuperscript{28} IGCM once occurred 10 days after surgical resection of thymoma.\textsuperscript{35} It has been postulated in the absence of direct evidence that alterations in immune function associated with thymoma predispose to giant cell myocarditis.\textsuperscript{22} Associations with sarcoma arising from the thymus\textsuperscript{13} and lymphoma\textsuperscript{36} have also been described; however, lymphoproliferative disease of the heart may be difficult to distinguish from giant cell myocarditis if myocyte necrosis is present.

The incidence of giant cell myocarditis is low and varies with the population studied and method of diagnosis. In a Japanese autopsy registry, the incidence of giant cell myocarditis was 0.007% (25 of 377,841 cases from 1958 to 1977).\textsuperscript{53} The incidence of giant cell myocarditis was a similarly low 3 of 12,815 necropsies from 1950 to 1963 at Oxford Infirmary.\textsuperscript{11} The incidence is higher when all IGCM cases diagnosed by endomyocardial biopsy or other surgical specimens, explanted heart, and autopsy are included. Using all pathologic specimens, 5 large, heart failure referral centers in the United States encountered a case of giant cell myocarditis about every 21 months from 1993 to 1997 (Giant Cell Myocarditis Registry, unpublished data).

Until recently, no easily available source of authoritative information on IGCM has been maintained. In 2001, the Web site www.gcminfo.org was established to provide such a resource for affected individuals and their families. This site also provides links to other relevant sites concerned with inflammatory heart disease.

**MECHANISMS OF GIANT CELL MYOCARDITIS**

The cause of human giant cell myocarditis is not known but often presumed to be autoimmune. Useful data supporting an autoimmune mechanism come from the Lewis rat model of IGCM. Experimental IGCM can be produced in the Lewis rat by autoimmunization with myosin.\textsuperscript{54} In this model, autoimmune giant cell myocarditis is mediated by CD4\textsuperscript{+} T cells that produce interferon-\gamma and macrophages that produce tumor necrosis factor and nitric oxide.\textsuperscript{55} The disease can be transferred by T lymphocytes.\textsuperscript{56} The histologic changes and hemodynamic deterioration are associated with inducible nitric oxide synthetase expression and attenuated by aminoguanidine, an inhibitor of inducible nitric oxide synthetase.\textsuperscript{57} The detrimental effect of nitric oxide in this model contrasts with the
beneficial effects of nitric oxide in coxsackie viral myocarditis.⁵⁸ Also of interest, tumor necrosis factor-alpha stimulates multinucleation of macrophages into giant cells in vitro when it is secreted from macrophages, but not when it is added exogenously.⁵⁹

Strain differences in rats with experimental giant cell myocarditis suggest that genetic factors play a major role in susceptibility to disease. Shioji and colleagues⁶⁰ reported the incidence, histopathology, and histocompatibility characteristics of 5 inbred strains of rats in which myocarditis was induced with porcine cardiac myosin. Immune-mediated myocarditis was induced in Lewis, Dahl (DIR/Eis) (RT-1), and Fisher rats but not in brown Norway rats or a second strain of Dahl rats (DIS/Eis) (RT-1). The disease was most severe in the Lewis rats and seemed to correlate with major histocompatibility complex class II region differences between the strains. Although 90% of cases in the Giant Cell Myocarditis Registry occurred in whites, firm conclusions regarding ethnic susceptibility cannot be drawn because these data may reflect the populations at the referral centers.¹⁵

Observations in human tissue suggest that IGCM is mediated by T lymphocytes as well.¹² In studies of paraffin-embedded tissue, infiltrating lymphocytes are almost always positive for T-cell antigens and giant cells are positive for macrophage antigens.²⁶,⁶¹,⁶² The T-cell subsets (helper and suppressor ratios) may vary during the evolution of the infiltrate. Electron microscopy has failed to find viral particles or other clues to the etiology of giant cell myocarditis.⁶²-⁶⁴

Data from the rat model of IGCM support aggressive immunosuppressive treatment early in the disease. Data from the rat model suggest that anti-T-lymphocyte antibodies and cyclosporine, but not prednisolone alone, prevent giant cell myocarditis.⁶⁵-⁶⁷ From these observations, immunosuppression with muromonab-CD3 and cyclosporine would be a reasonable treatment for patients who have giant cell myocarditis.

**DIAGNOSTIC STRATEGY**

The diagnosis of giant cell myocarditis should be considered for all patients with subacute heart failure of unknown cause. Of the 63 Multicenter Giant Cell Myocarditis Registry patients,¹⁵ 75% presented with congestive heart failure, 14% presented with ventricular arrhythmia, and lesser percentages presented with a syndrome mimicking acute myocardial infarction, heart block, or arterial embolization. The median time from symptom onset to presentation was 3 weeks. The median age was 42 years (range, 15 to 69 years), but patients younger than 15 years⁶⁸ and older than 70 years¹¹ have been reported. Men and women are affected equally, and cases have been described in many ethnic groups.

Common causes for heart failure and arrhythmia ought to be excluded per standard clinical practice. After a complete history, physical examination, electrocardiogram,
chest radiograph, an echocardiogram is usually done to exclude valvular and pericardial disease and cardiac masses. There are no specific echocardiographic findings to distinguish giant cell myocarditis from other forms of myocarditis, although the rapid decline in ejection fraction that may occur over several days in giant cell myocarditis patients is uncommon in lymphocytic myocarditis or cardiac sarcoidosis. Coronary angiography is superior to noninvasive stress imaging to exclude significant coronary stenosis or dissection. Magnetic resonance imaging or computed tomography may be done if clinically indicated to help exclude such disorders as arrhythmogenic right ventricular dysplasia or constrictive pericarditis.

Endomyocardial biopsy ought to be considered for patients with heart failure or ventricular arrhythmia of less than 3 months' duration who fail to improve despite optimal medical care. In most cases of lymphocytic myocarditis, the left ventricular ejection fraction improves with usual care, whereas the ejection fraction in giant cell myocarditis rarely improves. The development of ventricular tachycardia or heart block further increases the likelihood of giant cell myocarditis. The presence of associated disorders such as thymoma, myasthenia gravis, myositis, or inflammatory bowel disease (Table 17-3) may provide valuable clues as well.

Because giant cell myocarditis usually affects the endocardium, right ventricular endomyocardial biopsy may have a high sensitivity. In a substudy analysis of Giant Cell Myocarditis Registry subjects, Shields et al. found that endomyocardial biopsy had 82% to 85% sensitivity for giant cell myocarditis compared to the standard of surgical pathology (autopsy, explanted heart, or apical wedge section). This compares favorably to the roughly 35% sensitivity of endomyocardial biopsy in lymphocytic myocarditis, a more common but generally less severe and widespread process. However, the Shields et al. study included only subjects who had both endomyocardial biopsy and surgical specimens (ie, selected those with a particularly poor prognosis). The sensitivity of endomyocardial biopsy would likely be lower in an unselected population of heart failure subjects with giant cell myocarditis.

Sampling error is a concern in endomyocardial biopsy, and a minimum of 5 and sometimes more specimens ought to be obtained. Occasionally, the diagnostic lesion is seen only on additional cuts of the specimen blocks. Care must be taken to exclude hypersensitivity myocarditis, granulomatous myocarditis, foreign body reaction, and potential infectious causes by using standard diagnostic criteria and appropriate special stains. Once the diagnosis of giant cell myocarditis is certain, then one can consider the use of immunosuppressive agents in addition to usual care.

Other diagnostic techniques that have been suggested for viral myocarditis or cardiac sarcoidosis have not been applied to giant cell myocarditis. For example, antibodies to cardiac myosin have been described for patients with acute and chronic myocarditis.
Adenoviral and enteroviral DNA have been found in the hearts of patients with viral myocarditis. Magnetic resonance imaging and newer echocardiographic techniques have been applied for myocarditis and cardiac sarcoidosis in pilot studies. Nuclear imaging with gallium-67 to detect leukocytes or antimyosin antibodies to detect myocyte necrosis has not been used systematically to diagnose giant cell myocarditis. Because of the rarity and severity of giant cell myocarditis, a highly specific noninvasive test would be of great value; however, such a development is unlikely without a much greater understanding of the cause of giant cell myocarditis.

**TREATMENT OF GIANT CELL MYOCARDITIS**

Giant cell myocarditis is rapidly progressive and frequently requires the concurrent management of congestive heart failure, tachyarrhythmias, heart block, and secondary renal and hepatic insufficiency. Supportive care may include standard pharmacologic therapy for congestive heart failure, a permanent or temporary pacemaker, an implantable cardiac defibrillator, and an intra-aortic balloon pump. The use of these drugs and devices should be dictated by standard clinical practice.

Ventricular-assist devices have been used to bridge the time until patients with giant cell myocarditis have heart transplantation. Ventricular-assist devices have been used successfully as a bridge to transplantation in patients with lymphocytic or nonspecific myocarditis. Brilakis et al. reported a series of 9 patients from the Giant Cell Myocarditis Registry, who received ventricular-assist devices. Successful bridging to transplantation in 7 of 9 (78%) is similar to that reported for other recipients of assist devices. Post-transplantation survival of 57% (4 of 7) at 30 days and 29% (2 of 7) at 1 year was unexpectedly low. Poor post-transplantation survival may have been due to poor pre-transplantation condition of the patients. For a patient with giant cell myocarditis, the time to recovery has been bridged successfully with a biventricular Abiomed assist device (personal communication from patient).

Several case reports and the Giant Cell Myocarditis Registry suggest that treatment with certain combinations of immunosuppressants, but not steroids alone, prolongs transplant-free survival. The median time to death or cardiac transplantation for all 63 Registry subjects was 5.5 months from onset of symptoms. Seventy percent of affected individuals died or required heart transplantation within 1 year, and the overall rate of death or cardiac transplantation was 89%. Treatment with cyclosporine and steroid sometimes combined with azathioprine or muromonab-CD3 was associated with a median survival of 12.6 months compared with 3.0 months for those not treated with immunosuppressive agents ($P = 0.001$ by log-rank test). No published data exist for the use of
other immunosuppressive agents such as immunoglobulin, cyclophosphamide, tacrolimus, mycophenolate, or antithymocyte globulin for giant cell myocarditis.

These immunosuppressive treatment data must be interpreted cautiously. The data stem from a small registry, not a randomized controlled trial. They are subject to uncontrolled factors that could possibly lead to an observed treatment effect substantially larger than the actual treatment effect. As an example, it is possible that the giant cell myocarditis patients in this registry with longer transplant-free survival times were more likely to receive combined immunosuppression therapy than the patients with shorter times. If so, this would lead to a biased overestimate of the treatment effect.

The risks of aggressive immunosuppression in this setting are considerable. Cyclosporine can cause renal insufficiency, hypertension, liver function abnormalities, hirsutism, and gum enlargement. Muromonab-CD3 can cause profound hypotension, fever, chills, diarrhea, nausea, and vomiting. Long-term use of prednisone can cause osteoporosis and fractures, myopathy, cataracts, and glaucoma. Therefore, these drugs should be used to treat giant cell myocarditis by individuals experienced in their use at specialized centers.

Cardiac transplantation has been used with acceptable morbidity and mortality as a primary therapy for the management of giant cell myocarditis. Enthusiasm for transplantation was tempered by several reports of post-transplantation recurrence of giant cell myocarditis (Fig. 17-1). The 39 Giant Cell Myocarditis Registry patients who underwent heart transplantation had a 71% 5-year survival, despite a 25% post-transplantation histologic recurrence rate on surveillance endomyocardial biopsies. Therefore, overall post-transplantation survival for giant cell myocarditis patients is comparable to survival for patients who receive transplants for cardiomyopathy.

THE GIANT CELL MYOCARDITIS TREATMENT TRIAL

To control for possible bias in the Registry survival data and to investigate the mechanisms of giant cell myocarditis, the multicenter Giant Cell Myocarditis Treatment Trial was organized. This study is a randomized, open-label trial of muromonab-CD3, cyclosporine, and steroids (prednisolone followed by prednisone) versus cyclosporine and steroids for giant cell myocarditis diagnosed by endomyocardial biopsy. The primary efficacy end point is to compare the rate of death, transplantation, and ventricular-assist device placement at 1 year (event-free survival) in the 2 groups. To investigate the mechanism of survival benefit, hemodynamic and immunohistologic assessments will be obtained before treatment and during the study. The secondary efficacy end points include 1) change in left ventricular ejection fraction measured by radionuclide angiography, 2) improvement in myocardial inflammatory infiltrate, and 3) functional status assessed by the Living With Heart Failure
Fig. 17-1. Post-transplantation recurrence of giant cell myocarditis. (Courtesy of Edwina Duheing, MBBS, FRCPA.)

Questionnaire before and after 4 weeks of treatment. Adverse events will be monitored and assessed by an independent safety monitoring committee.

CONCLUSIONS

The prognosis of patients who have giant cell myocarditis is poor, but prolonged transplant-free survival may be possible with aggressive immunosuppression if treatment is started within several months of symptom onset. Because of possible bias in the retrospective Giant Cell Myocarditis Registry survival data and the substantial risks of treatment, the benefits of immunosuppression need to be confirmed in a prospective randomized trial. Consider the diagnosis of giant cell myocarditis for patients with less than 3 months of progressive congestive heart failure or ventricular arrhythmia. Patients with suspected IGCM might be referred to a center involved in the Giant Cell Myocarditis Treatment Trial. Despite a substantial rate of post-transplantation giant cell infiltration on surveillance biopsies, the post-transplantation survival of giant cell myocarditis patients is comparable to post-transplantation survival for cardiomyopathy patients.
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Myocarditis: From Bench to Bedside


Myocarditis: From Bench to Bedside


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